



Freshwater algae of Laknavaram Lake from Warangal District, Telengana State, India

L.Dup Singh, N.Ella Swamy, B.Kumara Swamy and B.Digamber Rao

Department of Botany, Kakatiya University, Warangal, Telangana State, India-506 009, Mail: bdr_kuc@yahoo.co.in

ABSTRACT

The Laknavaram Lake of Warangal District, Telangana State is an important Eco-tourist place, with beautiful rocky hills built by the mighty Kakatiya kings, during the period of 12th century and these water bodies are surrounded by thick forest with rocky hills. The lake Laknavaram is the main source of irrigation and drinking water and it was constructed by Sri Ganapathy Rudradeva of Kakatiya dynasty in the year 1213AD. The lake water body being used for agriculture, drinking and other recreational purposes (swimming, bathing, fishing and boating), an attempt has been made in the study to correlate the periodicity of phytoplanktons in the lake with physico-chemical parameters of the water body under study. The lake under study are important component of water body exposed to varying degrees of cultural eutrophication to some extent and are therefore regarded as more suitable production studies in the scheme of its reclamation. Since the study was undertaken over a period of two years (Sept. 2010-Aug. 2012) and the changes in chemical composition of water and algal populations were recorded at regular intervals, it may be presumed that the data gives a reasonably accurate basis for the interpretation of the distribution and periodicity of phytoplankton of the waters. In view of this the present study was undertaken to elucidate certain aspects of phytoplanktons of selected lake of Warangal district (Telangana State) with regard to its water quality.

INTRODUCTION

Algae are a large and diverse group of simple, typically autotrophic organisms, ranging from unicellular to multicellular forms. In aquatic ecosystems phytoplankton play an important role in the ecology of water bodies through primary production. Studies on planktonic composition and physico-chemical characteristics of water are necessary to acquire basic knowledge on the biodiversity status of a water body. Algal flora varies from season to season and an important feature of freshwater algal flora is its cosmopolitanism. The phytoplanktonic study is a very useful tool for the assessment of water quality in any type of water quality in any type of water body and also contributes to understanding of the basic nature of lakes (Pawar et al. 2006). The necessity of using phytoplankton as effective and appropriate method of biomonitoring for evaluation of water quality has been emphasized (Annalakshmi and Amsath, 2012). Considerable work has been done in India about systematic survey, distribution, periodicity and ecology of algae in different habitats (Das et al., 2009, Adhikary et al. 2010, Das et al., 2010, Bhakta et al., 2010, Das and Adhikary, 2012a, b & c and Kumaraswamy et al. 2013). Recently Mahajan (2005), Jayakumar and Karpaganu (2005), Sanap et al., (2006), Shoba et al., (2006), Harsha Sadanand et al., (2006) Mruthunjaya and Rajendra Prasad (2006), Mallishwar et al., 2007, Gupta and Anuj Bhadauriya 2007, Roy Zacharias and Joy (2007), Maya Subramoni, 2007, Kavitha and Rajini Balasingh (2007), Mruthunjaya et al., 2007. Reshma and Prakasam (2007), Girija Kumari and Abraham (2007), Sanap et al., (2008), Mista et al., (2008). Murugesan and Shivasubramanian, (2008), Aarti Narsimhan et al. (2008), Mista et al., (2009), Latha and Ramachandra Mohan (2010), Leela et al., (2010), Ramadosu and Sivakumar (2010) and Chinnaiah et al., (2011) studied on various fresh water bodies and described about physico-chemical characteristics along with algal population studies. Hence, the present study was undertaken to know the influence of physico-chemical parameters of water on algal populations and their seasonal changes of Laknavaram lake.

MATERIAL AND METHODS

The water samples collected from the lake in bottles were brought to the laboratory for analysis as per the standard methods described by APHA (1985) and Trivedy and Goel (1986). Four sampling sites were identified almost equidistant on the shore of the lakes to its north, east, west and south. The phytoplanktonic forms were collected by plankton net No. 20 silk bolting cloth. Sometimes samples for quantitative and qualitative analysis of phytoplankton was collected in 1 litre capacity. Pyrex glass bottles where 1ml of Lugol's solution was kept prior to the filling of the bottle by lakes water from below the surface. For sedimentation

of plankton material the bottle was kept undisturbed in the dark for 48 hrs at room temperature. After that the overlying water from the bottle was decanted and the final volume was adjusted in between 10 to 15ml. The latter method was usually used to compare whether there was any demerit of using the other method. No significant difference was observed. After collection the phytoplankton material was transferred to glass preserved permanently in Transeau's solution (Distilled water 60ml or 6 parts; Absolute alcohol 30ml or 3 parts; Formaldehyde 10 ml or 1 part. To each 100 ml of the above solution 5ml of glycerine was added to prevent the materials from becoming brittle (Transeau, 1951).

Mounting Fluid:

Ten percent glycerine solution in distilled water was used as mounting fluid for the preparation of temporary and semi permanent slides for microscopic study. Cotton blue and Lacto- phenol were used as mounting fluid and as stain, which facilitated a clean view of the materials under the microscope.

Phytoplanktons were identified by using standard key books (Patrick and Reimer, 1966, Suxena and Venkateshwarlu, 1968; Philipose, 1967, Desikachary, 1959; Prescott, 1951 and Smith, 1933).

Phytoplankton analysis:

The qualitative enumeration of the phytoplankton were carried out with the help of a Sedgwick Rafter counting cell and by drop method.

The counting cell was filled with water to test for any leakage and it was emptied and dried properly. The vials containing the concentrate were shaken properly and 1ml of concentrated sample taken in to the Sedgwick Rafter counting cell. Numerical counts of all members of phytoplanktons were made using the Whipple micrometer. The organisms thus counted, were expressed as units/liter (units/l) of the sample. Seasonal variation and abundance of the dominant algal genera and the groups of phytoplankton were noted.

The abundance of phytoplankton groups were calculated according to the following formulae (Welch, 1948).

$$N = \frac{(a \times 100)C}{L}$$

N = Number of phytoplanktons per liter of original water

a = Average number of phytoplankton in all counts in the counting cells

C = Volume of original concentrate in ml.

L = Volume of water passed through the net.

The result was expressed as unit/liter

RESULT AND DISCUSSIONS

The water temperature varied from 23°C to 31°C. Turbidity of the water ranged from 0.03 to 8.00 NTU, conductivity annual average value varied from 0.74 to 0.89 milli mhos/cm in 2010-2011 and from 0.46 to 0.62 m mhos/cm in 2011-2012. Transparency of water ranged from 18 to 133 cm, TDS from 160.00 to 680.00 mg/l, TSS varied from 160.40 to 580.80 mg/l. The bottom substrata of the lake was of sandy loamy in nature. The pH values varied from 7.0 to 8.4 in two years of study period, while the DO values ranged from 6.5mg/l to 10.0 mg/l. In Primary productivity the GPP values ranged from 0.0045 to 3.6105 mg c/hr/l,

the NPP varied from 0000 to 3.56 185 mg c/hr/l and Resp^N ranged from 0000 to 2.7445 mg c/hr/l. The oxidation-reduction potential (E_h) was shown variation between 0.211 to 0.372, Oxidation-reduction index values ranged from 24.80 to 2745. The Biochemical oxygen demand fluctuated between 3.0 to 21.0 mg/l and the Chemical oxygen demand varied from 0.32 to 0.95mg/l respectively. Silicates, free CO₂. Phosphates, Total Hardness and Chloride, values ranged from 0.0002 to 0.0084mg/l; 4.00 to 47.00 mg/l; 0.04 to 0.82 mg/l; 110 to 330 mg/l and 10.0 to 125.0 mg/l. In total alkalinity the OH alkalinity was recorded only in four months during the period of study and its values varied from 10 to 60mg/l, the phenolphthalein alkalinity ranged from 20 to 300 mg/l and the alkalinity varied from 10 to 260 mg/l. No trace of nitrate, nitrite and ammonia were observed. The monthly analysis of Chromium, Cadmium, Cobalt, Lead, Mercury and Zinc were undertaken and no trace of heavy metals could be detected in the present investigation.

The algal members comprised of 63 genera of which 14 belonged to Bacillariophyceae, 27 to Chlorophyceae, 17 to Cyanophyceae and 5 to Euglenophyceae (Table-1). Highest number of individual was represented by green algae (452782) followed by blue- green algae (405130), diatoms (1113908) and the members of Euglenophyceae (32167). A total of 76 forms of phytoplanktons were recorded. The total percentage of phytoplankton community were 78.15% and 21.85% are others. The seasonal abundance of the Laknavaram lake have shown the two most prominent blue greens, *Gloeotrichia natans*, *Lyngbya majuscula*, two diatoms, *Bacillaria paradosa*, *Pinnularia viridis*. and green alga, *Pithophora varia*. The compound index of Nygaard algal indices have shown the results in which they varied from 1.4 to 5.8 with the average value of 3.67 under investigation.

Seasonal abundance of the prominent phytoplanktons of Laknavaram lake has also been studied in which the two most prominent blue green algae *Gloeotrichia natans*, *Lyngbya majuscula* two important diatoms, *Pinnularia viridis*, *Bacillaria paradosa*. and green-alga *Pithophora varia* in the year 2010-2011. Of the two most prominent Cyanophyceae members *G.natans* was found to be dominant in March 8.0 units 1⁻¹ X 10⁴ and minimum in January 2.0 units 1⁻¹ X 10⁴. *Lyngbya majuscula* has been shown to be dominant in February 5.4 units 1⁻¹ X 10⁴ and lowest in October 0.2 units 1⁻¹ X 10⁴. The Bacillariophyceae members of *Bacillaria paradosa*, was observed more dominantly in the month of March 8.0 units 1⁻¹ X 10⁴ and the lowest in the month of September 1.0 units 1⁻¹ X 10⁴. *P.viridis*, was maximum in March 7.2 units 1⁻¹ X 10⁴, with minimum in September 3.2 units 1⁻¹ X 10⁴. The green algae, *Pithophora varia* was the only genus observed most dominantly in July 4.9 units 1⁻¹ X 10⁴ and least in 0.4 units 1⁻¹ X 10⁴ in the month of October under study. During the observation period between September 2011 - August 2012, the seasonal abundance of most prominent phytoplanktons were exhibited with two members of Cyanophyceae (*Gloeotrichia natans* and *Lyngbya majuscula*), two members of diatoms (*Bacillaria paradosa*, and *Pinnularia viridis*), and one genus of *Chlorophyceae* member, *Pithophora varia*. *Gloeotrichia natans* was maximum in March 6.4 units 1⁻¹ X 10⁴ and minimum in August 1.8 units 1⁻¹ X 10⁴, *Lyngbya majuscula* was observed dominantly in the month of February 6.0 units 1⁻¹ X 10⁴ and lowest in August 0.8 units 1⁻¹ X 10⁴. The diatom member *Bacillaria paradosa*., was found to be maximum in March, 9.8 units 1⁻¹ X 10⁴ and minimum in August 1.9 units 1⁻¹ X 10⁴, *Pinnularia viridis* was found more in March 7.8 units 1⁻¹ X 10⁴ and less in August 1.9 units 1⁻¹ X 10⁴. Of the one prominent green alga available *Pithophora varia* was found maximum in August 5.0 units 1⁻¹ X 10⁴ and minimum in April 0.5 units 1⁻¹ X 10⁴. All the members of Cyanophyceae, Bacillariophyceae and Chlorophyceae were found in almost all the seasons and in every month of 2011-2012. The phytoplankton populations of the lake varied with the seasonal variations and the maximum phytoplankton production coincided with the optimum water depth of one meter under study.

Table-1, Phytoplankton species found in the Laknavaram Lake

Sl. No.	BACILLARIOPHYCEAE (14 forms) 14 genera
1)	<i>Actinastrum sp.</i>
2)	<i>Amphora sp.</i>
3)	<i>Bacillaria paradosa</i>
4)	<i>Cocconeis sp.</i>
5)	<i>Diatoma sp.</i>
6)	<i>Gomphonema gracile</i>
7)	<i>Gyrosigma sp.</i>
8)	<i>Navicula exigna</i>
9)	<i>Nitzschia sp.</i>
10)	<i>Pinnularia viridis</i>
11)	<i>Tabellaria sp.</i>
12)	<i>Gyrosigma scalproides</i>
13)	<i>Neidium gracile</i>

14)	<i>Diademesmis confervacea</i>	41)	<i>Chara vulgaris</i>
	EUGLENOPHYCEAE(5 forms)	42)	<i>Chlorella vulgaris</i>
	5 genera	43)	<i>Closterium tumidum</i>
15)	<i>Euglena caudata</i>	44)	<i>Cl. porrectum</i>
16)	<i>Phacus sp.</i>	45)	<i>Cladophora glomarata</i>
17)	<i>Trachelomonas curta</i>	46)	<i>Cl. crispata</i>
18)	<i>Menoidium sp.</i>	47)	<i>Cosmarium botrytis</i>
19)	<i>Lepocinalis fusiformis</i>	48)	<i>C. auriculatum</i>
	CYANOPHYCEAE (19 forms)	49)	<i>C. granatum</i>
	17 genera	50)	<i>Euastrum verrucosum</i>
20)	<i>Anabaena iyengarii</i>	51)	<i>Microspora sp.</i>
21)	<i>Anabaenopsis sp.</i>	52)	<i>Mougeotia sp.</i>
22)	<i>Aphanocapsa litorales</i>	53)	<i>Oedogonium borisianum</i>
		54)	<i>O. grande</i>
23)	<i>Arthrospira sp.</i>	55)	<i>Oocystis gigas</i>
24)	<i>Chroococcus minutus</i>	56)	<i>Pediastrum duplex</i>
25)	<i>Gloeocapsa atrata</i>	57)	<i>Pithophora varia</i>
26)	<i>Gloeotrichia natans</i>	58)	<i>Protococcus sp.</i>
27)	<i>Hydrococcus sp.</i>	59)	<i>Rhizoclonium hieroglyphicum</i>
28)	<i>Microcystis aeruginosa</i>	60)	<i>Scenedesmus denticulatus</i>
29)	<i>Nostoc sphaerium</i>	61)	<i>S. quadricauda</i>
30)	<i>Oscillatoria formosa</i>	62)	<i>S. dimorphus</i>
31)	<i>O. tenuis</i>	63)	<i>S. obliquus</i>
32)	<i>O. rubescens</i>	64)	<i>Spirogyra acanthospora</i>
33)	<i>Phormidium molle</i>	65)	<i>S. discoidea</i>
34)	<i>Synechococcus sp.</i>	66)	<i>S. formosa</i>
35)	<i>Trichodesmium sp.</i>	67)	<i>Stigeoclonium tenue</i>
36)	<i>Lyngbya ceylanica</i>	68)	<i>Staurostrum pinnatum</i>
37)	<i>Planktothrix compressa</i>	69)	<i>Tetraedron quadratum</i>
38)	<i>Homoeothrix janthiana</i>	70)	<i>Ulothrix sp.</i>
	CHLOROPHYCEAE (38 forms)	71)	<i>Zygnema czurde</i>
	27 genera	72)	<i>Z. areolatum</i>
39)	<i>Ankistrodesmus falcatus</i>	73)	<i>Zygnemopsis sp.</i>
40)	<i>Bulbochaete sp.</i>	74)	<i>Radiococcus nimbatus</i>
		75)	<i>Characium ambiguum</i>
		76)	<i>Anthrodesmus curvatus</i>

BACILLARIOPHYCEAE:

Among the recorded 14 genera with 14 forms of diatoms only a few (*Actinastrum sp.*, *Amphora sp.*, *Bacillaria paradosa*, *Cocconeis sp.*, *Diatoma sp.*, *Diadesmus confervacea*, *Gomphonema gracile.*, *Gyrosigma sp.*, *Gyrosigma scalproides*, *Navicula exigna*, *Neidium gracile*, *Nitzschia sp.*, *Pinnularia viridis*, *Tabellaria sp.*) was found to be well adapted to the concerned habitats of the lake. Throughout the period of study the water body of the lake have showed the highest abundance of diatom (1113908 cells/l) under investigation. Among the recorded genus of diatom species diversity was exhibited by the genus *Gyrosigma sp.*, (2 sp.) respectively. (Table-1).

CHLOROPHYCEAE:

The generic diversity of Chlorophycean members were maximum with 27 genera with 38 forms during the period of study in the lake under investigations (**Table-1**). Such as *Ankistrodesmus falcatus*, *Anthrodesmus curvatus*, *Bulbochaete sp.*, *Chara vulgaris*, *Chlorella vulgaris*, *Characium ambiguum*, *Closterium tumidum*, *Cl.porrectum*, *Cladophora glomarata*, *Cl. crispata*, *Cosmarium botrytis*, *C.auriculatum*, *C.granatum*, *Euastrum verrucosum*, *Microspora sp.*, *Mougeotia sp.*, *Oedogonium borisianum*, *O.grande*, *Oocystis gigas*, *Pediastrum duplex*, *Pithophora varia*, *Protococcus sp.*, *Radiococcus nimbatus*, *Rhizoclonium hieroglyphicum*, *Scenedesmus denticulatus*, *S.quadricauda*, *S. dimorphus*, *S. obliquus*, *Spirogyra acanthospora*, *S.discoidea*, *S. formosa*, *Stigeoclonium tenue*, *Staurastrum pinnatum*, *Tetraedron quadratum*, *Ulothrix sp.*, *Zygnema czurde*, *Z. areolatum*, and *Zygnemopsis sp.*, were recorded as the major dominant genus in the sampling lake under investigation.. Throughout the period of study the water body have shown the highest recorded abundance (452782 cells/l) of green algae, *Scenedesmus* was the genus which exhibited the maximum (4 species) diversity followed by , *Spirogyra* (3 species), *Cosmerium sp.* (3) *Closterium* (2 species) and *Oedogonium*, *Pediastrum*, *Tetraedron* and *Zygnema* (2 species) respectively. Throughout the sampling period peaks of green algae were recorded in monsoon months and lower abundance in summer months.

CYANOPHYCEAE:

Throughout the period of study sharp peaks of Cyanophycean members were exhibited in summer months and they exhibited with 17 genera with 19 forms only a few of them were found to be the dominant in the sample lake, these are *Anabaena iyengarii*, *Anabaenopsis sp.*, *Aphanocapsa littorales*, *Arthrospira sp.*, *Chroococcus minutes*, *Gloeocapsa atrata*, *Gloeotrichia natans*, *Hydrococcus sp.*, *Microcystis aeruginosa*, *Nostoc sphaerium*, *Oscillatoria formosa*, *O.tenuis*, *O.rubescens*, *Phormidium molle*, *Synechococcus sp.*, *Trichodesmium sp.*, and *Lyngbya ceylanica*, *Planktothrix compressa* and *Homoeothrix janthiana*, respectively. Blooms of cyanophycean algae were recorded during the summer season. The blooms of *Microcystis aeruginosa* and *Lyngbya ceylanica* were reported from Laknavaram lake. During the period of study the lake was expressed with higher abundance of blue-green algae under observation (405130 cells/l). Among the observed members of Cyanophyceae maximum species diversity was exhibited by the *Oscillatoria* (3 species) only.

EUGLENOPHYCEAE:

The recorded 5 genera with 5 forms in Euglenophyceae (*Euglena caudata*, *Phacus sp.*, *Lepocinclis fusiformis*, *Menoidium sp.* and *Trachelomonas curta*) were found to be well adapted to the concerned habitats under study. The Laknavaram lake showed abundance of Euglenoides (32167 cells/l) with large peaks of Euglenoid population were reported in winter months, while small peaks were found in summer months. No species diversity was found the Euglenoids under study.

The average percentage of algae during the period of Sep. 2010-Aug 2012:

In the lake of Laknavaram, the phytoplankton community represents the presence of 30.49% of Bacillariophyceae, 42.59% of Chlorophyceae, 24.27% of Cyanophyceae and 2.65% of Euglenophyceae. The Chlorophyceae members were dominant over other members.

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